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Two new polyoxygenated cyclohexenes from *Uvaria kweichowensis*

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Two new polyoxygenated cyclohexenes, kweichowenols C and D, were isolated from the leaves of *Uvaria kweichowensis*, and their structures were established on the basis of their spectral data. The two new compounds showed some antitumor activity by the MTT assay.

Keywords: Annonaceae; *Uvaria kweichowensis*; leaves; cyclohexenes; kweichowenol C; kweichowenol D

1. Introduction

During the past two decades, the genus *Uvaria* has been found to be a rich source of polyoxygenated cyclohexenes, some of which exhibited antitumor, antileukemic, or anti-biotic activities [1,2]. Our previous studies have resulted in the isolation of a series of polyoxygenated cyclohexenes from *Uvaria grandiflora* and *Uvaria tonkinensis* var. *subglabra* [3,4]. *Uvaria kweichowensis* is an herb used to cure inflammation and tumor in the southwest area of China. During the course of our investigation for antitumor agents from the chloroform extract of the leaves of *U. kweichowensis*, besides kweichowenols A [5] and B [5], two new crystalline compounds, kweichowenols C (**1**) and D (**2**) were obtained. We now report the isolation and structural elucidation of the two new compounds, as well as the evaluation of their antitumor activities.

2. Results and discussion

Kweichowenol C (**1**) was obtained as white needles. The molecular formula was determined to be C₂₃H₂₂O₇ by HR-ESI-MS, which

showed quasi-molecular ion peaks at m/z 411.1436 [M + H]⁺, 433.1267 [M + Na]⁺, and 449.1012 [M + K]⁺. The IR spectrum of compound **1** suggested the presence of hydroxyl group(s) (3587, 3454, 1115, and 1090 cm⁻¹), ester group(s) (strong absorption at 1722 cm⁻¹) and monosubstituted phenyl ring(s) (1601, 1454, and 710 cm⁻¹). The UV spectrum gave absorption maxima at λ 230, 274, and 280 nm, indicating the presence of one or two benzoyl groups.

The most informative evidence for the structural elucidation of compound **1** was derived from its ¹H NMR spectral data, which, together with those of other related compounds, are summarized in Table 1. The aromatic proton signals between δ 7.3 and 8.1 (10H, m) confirmed the presence of two benzoyl moieties, while the ethidine moiety in the structure was indicated by the proton signals at δ 1.24 (3H, d) and 5.26 (1H, q). The ¹H NMR spectrum also showed two olefinic proton signals at δ 5.87 (2H, m) and a carbonyl proton signal (secondary alcohol) at δ 4.40. The two benzoyl groups appeared to be located at C-3 and C-6 based upon the downfield shifts of H-3 at δ 5.59 and H-6 at δ

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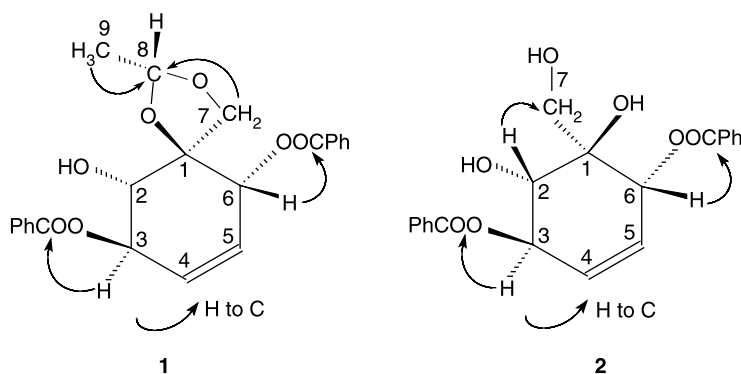
Table 1. ^1H (600 MHz) and ^{13}C (150 MHz) NMR spectral data of compounds **1** and **2** (CDCl_3 , δ ppm).

	1		2	
	δ_{C}	δ_{H}	δ_{C}	δ_{H}
1	84.8		75.6	
2	73.8	4.40 d (8.4) ^a	76.7	4.21 d (8.4)
3	74.1	5.59 dd (8.4, 2.4)	74.0	5.78 dd (8.4, 2.4)
4	129.6	5.87 m	128.6	5.85 m
5	127.4	5.87 m	127.3	5.87 m
6	73.3	5.84 d (2.4)	75.8	5.80 m
7	65.6	4.39 d, 4.26 d (8.4)	62.7	4.17 d, 4.11 d (12.0)
8	103.8	5.26 q (4.2)		
9	19.7	1.24 d (4.2)		
Ar	120–140	7.3–8.1 m	120–140	7.3–8.1 m
Carbonyl	166.0, 166.7		166.3, 166.7	

^aData in parentheses are J values (in Hz).

5.84, while the ethidine group was established at C-1 and C-7 on the basis of the downfield shifts of H-7 at δ 4.39 and 4.26. The elucidation of compound **1** was supported by the existence of the fragment ion peaks at m/z 294 and 116 resulted from the typical retro-Diels–Alder (RDA) cleavage of the molecule of compound **1** in the EI-MS spectrum. In comparison with the ^1H NMR spectral data of uvarigranol F [6], the coupling constant $J_{2,3}$ of 8.4 Hz suggested that H-2 and H-3 were favorably located at axial position. In addition, $J_{5,6}$ of 2.4 Hz suggested that the benzoxy group at C-6 was likely located at the pseudo-equatorial position [7]. The proposed structure of compound **1** was verified by the HMBC experiment

(Figure 1). The relative stereochemistry of compound **1** was further established from NOESY interaction. The interactions between H-3 (δ 5.59) and H-7 (δ 4.39 and 4.26) indicated that they were on the same side of the molecule, while that between H-2 (δ 4.40) and H-6 (δ 5.84) suggested that H-2 and H-6 were on the other side. The interactions between H-8 (δ 5.26) and H-2 (δ 4.40) indicated that the relative stereochemistry of the methyl located at C-8 was α -configuration. The CD spectrum of compound **1** exhibited a split curve centered at λ 227 nm with a positive Cotton effect at λ 235 (+4.62) and a negative Cotton effect at λ 219 (−9.86), indicating the clockwise manner for the orientation of the two benzoxy

Figure 1. Key HMBC correlations of compounds **1** and **2**.

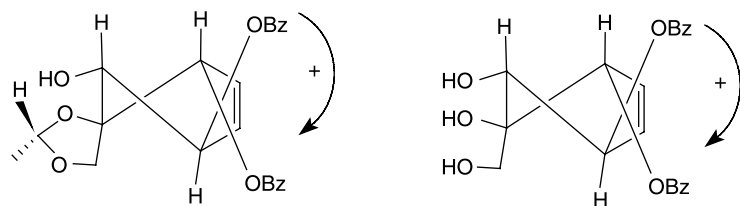


Figure 2. The chirality of the dibenzoates of compounds **1** and **2**.

groups at C-3 and C-6 (Figure 2); thus the absolute configuration was established as *1R*, *2S*, *3R*, *6R*, *8R* [8].

In order to determine whether compound **1** was naturally occurring in the leaves of *U. kweichowensis*, the leaves were extracted with CHCl_3 , and the extract was chromatographed over silica gel column, the fraction eluted by petroleum ether–EtOAc (85:15–82.5:17.5) was subjected to HPLC analysis. The HPLC experiment demonstrated the presence of kweichowenol C in the CHCl_3 extract of the leaves of *U. kweichowensis*. Such a result indicated that compound **1** occurred naturally, rather than as a consequence of the utilization of acetone during the separation procedure.

Kweichowenol D (**2**) was obtained as white solid. The molecular formula was determined to be $\text{C}_{21}\text{H}_{20}\text{O}_7$ by HR-ESI-MS, which showed quasi-molecular ion peaks at m/z 385.1284 $[\text{M} + \text{H}]^+$, 407.1067 $[\text{M} + \text{Na}]^+$, and 423.1067 $[\text{M} + \text{K}]^+$. Compound **2** has similar IR and UV spectra to those of compound **1**. Its ^1H NMR spectrum displayed the signals of two benzoate groups (δ 7.3–8.1, 10H, m) and seven protons on a methyl–cyclohexene skeleton (δ 4.2–5.9; the assignment shown in Table 1). Among the latter signals, an AB quartet at δ 4.17 and 4.11 (10.8 Hz) was attributed to the two methylene protons at C-7, similar to those of the known polyoxygenated cyclohexenes.

The HMBC spectrum of compound **2** (Figure 1) indicated that the signal at δ 4.21 was attributed a carbonyl proton at C-2 for the correlation between it with C-7 (δ 62.7). The correlations between H-3 (δ 5.78) and the

ester carbonyl at δ 166.7 and between H-6 (δ 5.80) and the ester carbonyl at δ 166.3 indicated that the two benzoate groups are located at C-3 and C-6, respectively.

Based upon a careful analysis of the coupling constants and the correlations shown in the NOESY spectrum, the relative stereochemistry of compound **2** was established. The $J_{2,3}$ value of 8.4 Hz revealed that both H-2 and H-3 are located at axial positions. In the NOESY spectrum of compound **2**, interactions between H-3 (δ 5.78) and H-7 (δ 4.17 and 4.11) indicated that they were on the same side of the molecule, while those between H-2 (δ 4.21) and H-6 (δ 5.80) suggested that H-2 and H-6 were on the other side.

The CD spectrum of compound **2** exhibited a positive Cotton effect at λ 235.5 (+3.55) and a negative Cotton effect at λ 218.5 (–6.16), indicating that the two benzoate groups were oriented in a clockwise manner. Since the overall relative configuration was known, the absolute configuration of compound **2** was established as *1R*, *2S*, *3R*, *6R*.

In order to determine whether compound **1** was an acetalation product of compound **2**, compound **1** was hydrolyzed by TFA/ H_2O [9], then the hydrolysis product was subjected to HPLC analysis. The HPLC experiment demonstrated the presence of compound **2**. Such a result indicated that compound **1** was an acetalation product of compound **2**.

In the case of bioactivities of compounds **1** and **2**, antitumor activities of them were expressed as IC_{50} values. As determined by MTT assay, the IC_{50} values of compounds **1** and **2** against A549 bronchogenic carcinoma cells were 55 and 22 $\mu\text{g}/\text{ml}$, respectively,

SK-MES-1 bronchogenic carcinoma cells 58 and 25 $\mu\text{g/ml}$, and NCI-H446 bronchogenic carcinoma cells 48 and 19 $\mu\text{g/ml}$. The results suggested that the antitumor activity of compound **2** was a little better than that of compound **1**.

3. Experimental

3.1 General experimental procedures

Melting points were determined using a Fisher–Johns melting point apparatus and are uncorrected. Optical rotations were obtained on a PerkinElmer model 241 polarimeter. UV spectra were measured on a Shimadzu UV-2401 spectrometer. IR spectra were taken on a PerkinElmer 983 G spectrometer. ^1H , ^{13}C NMR, and 2D-NMR spectra were recorded on a Varian Inova 600 spectrometer in CDCl_3 using tetramethylsilane as internal standard. EI-MS and FAB-MS spectra were determined on a Micromass Zabspec spectrometer. HR-ESI-MS spectra were determined on a Q-TOF2 spectrometer. CD spectra were determined on a JASCO-715 spectrometer. Preparative HPLC was carried out on a column of Phenyl (250×9.4 mm i.d., Agilent Zorbax SB-Phenyl; Palo Alto, CA, USA) with a Waters 2996 detector; the flow rate was 3 ml/min and the wave length for detection was 230 nm. MPLC was carried out on a column of silica gel H (460×26 mm i.d., Buchi Borosilikat 4.6; Flawil, Switzerland). Silica gel (200–300 mesh) for column chromatography was obtained from Qingdao Marine Chemical Factory, Qingdao, China. Precoated plates of silica gel GF₂₅₄ were used for TLC, and detected under UV.

3.2 Plant material

The leaves of *U. kweichowensis* were collected in Guizhou Province of China in November 2002, and identified by Prof. Shou-Quan Lin and Dr Guo-Qiang Li of our institute. A voucher sample is deposited in the Herbarium of the Institute of Medicinal Plant Development, Peking Union Medical College.

3.3 Extraction and isolation

The dried plant material (5 kg) was extracted three times with 95% EtOH under reflux. The solvent was subsequently dried under reduced pressure to give the residue, which was partitioned between CHCl_3 and H_2O . The CHCl_3 -soluble fraction was further portioned between petroleum ether and 90% MeOH (V/V). The 90% MeOH fraction (85 g) was chromatographed over silica gel column, which was eluted with petroleum ether–EtOAc gradients to afford 25 fractions (F1–F25). Further purification of F8 through MPLC, using petroleum ether–EtOAc (85:15) as eluent, yielded kweichowenol C (**1**; 15 mg). Kweichowenol D (**2**; 130 mg) was obtained from F15 through RP-HPLC using MeOH– H_2O (80:20) as eluent.

3.3.1 Kweichowenol C (**1**)

White needles, mp 48–49°C. $[\alpha]_{20}^{\text{D}}$ –92.6 ($c = 0.001$, CHCl_3). UV λ_{max} (CHCl_3 , nm): 230, 274, 280. IR (KBr; ν_{max} , cm^{-1}): 3587, 3454, 1722, 1601, 1454, 1273, 1115, 710. For ^1H and ^{13}C NMR (CHCl_3) spectral data, see Table 1. EI-MS m/z : 395 (6.0), 294 (4.5), 273 (6.5), 227 (6.5), 203 (9.0), 130 (6.5), 122 (18.6), 105 (100), 77 (35). FAB-MS m/z 411 ($[\text{M} + \text{H}]^+$). HR-ESI-MS m/z : 411.1436 ($[\text{M} + \text{H}]^+$ (calcd for $\text{C}_{23}\text{H}_{23}\text{O}_7$, 411.1444).

3.3.2 Kweichowenol D (**2**)

White solid, mp 63–64°C. $[\alpha]_{20}^{\text{D}}$ –85.2 ($c = 0.002$, CHCl_3). UV λ_{max} (CHCl_3 , nm): 230, 274, 280. IR (KBr; ν_{max} , cm^{-1}): 3438, 3261, 1728, 1701, 1599, 1583, 1452, 1273, 1115, 710. For ^1H and ^{13}C NMR (CHCl_3) spectral data, see Table 1. EI-MS m/z : 294 (7.5), 273 (9.0), 227 (5.5), 203 (11.5), 130 (8.5), 122 (25.5), 105 (100), 77 (45). FAB-MS m/z : 385 ($[\text{M} + \text{H}]^+$). HR-ESI-MS m/z : 385.1284 ($[\text{M} + \text{H}]^+$ (calcd for $\text{C}_{21}\text{H}_{21}\text{O}_7$, 385.1287).

3.4 Antitumor bioassays

To evaluate the antiproliferative effect of compounds **1** and **2** on A549, SK-MES-1, and

NCI-H446 bronchogenic carcinoma cell lines, the MTT colorimetric assay was performed. The amount of formazan was determined by a photometer at 570 nm. Cells were plated onto 96-well flat-bottomed culture plates at a concentration 5×10^4 cell per well in complete RPMI 1640 culture medium. Twenty-four hours after plating, the medium containing fetal calf serum was removed and test solutions were given to the cells in various final concentrations such as 10, 25, 50, 100, and 200 $\mu\text{g/ml}$. After incubation with drugs for 24 h, the MTT solution was added to the wells and plates were incubated at 37°C for 4 h. Results were expressed as percentage of the absorbance in control cells compared with those in the drug-treated cells.

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